Selective Fluorometric Sensing of Calcium Cation by C-Pivot Lariat Monoaza-crown Ether with Two Pyrene Moieties

Yohji Nakatsuji,* Misako Nakamura, Tomoya Oka, and Masahiro Muraoka Department of Applied Chemistry, Faculty of Engineering, Osaka Institute of Technology, 5-16-1 Omiya, Asahi-ku, Osaka 535-8585

(Received August 5, 2011; CL-110658; E-mail: nakatsuji@chem.oit.ac.jp)

A new type of C-pivot lariat monoaza-crown ether ${\bf 1b}$ containing a 15-crown-5 ring and two pyrene moieties on the sidearms was found to be a highly sensitive and selective fluorescent chemosensor toward ${\rm Ca^{2+}}$ in the presence of ${\rm Na^+}$, ${\rm K^+}$, ${\rm Mg^{2+}}$, and ${\rm Ca^{2+}}$.

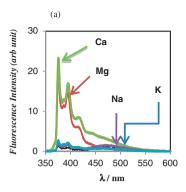
Much attention has been focused on the development of sensing techniques for specific cations in a variety of research fields of chemistry, biology, and medicine.1 Selective and sensitive chemosensors for Na⁺, K⁺, Mg²⁺, and Ca²⁺ are strongly desired for detection of these cations in living organisms.² Supramolecular chemosensors used for cation recognition are composed of a binding part for guest cations and a sensing part to read information about the complexation.³ Thus, a number of chemosensors have been developed by an elaborate combination of both binding and sensing parts.³ Recently, we found that C-pivot lariat crown ethers⁴ with two pyrenylmethyl groups on the electron-donating sidearms are potentially useful as selective chemosensors for alkaline earth metal cations.⁵ In this case, effective coordination of the electron-donating sidearm to metal cations resulted in changing the spatial distance of the two pyrene rings. The ratiometry of monomer and excimer emissions of the pyrene moieties⁶ was successfully used for the selective detection of metal cations. A combination of this sensing mechanism with photoinduced electron transfer (PET), another effective method for detecting metal cations,^{3,7} should afford an interesting strategy to develop new fluorescent chemosensors. From this point of view, we describe the design and synthesis of novel chemosensors derived from C-pivot lariat monoaza-crown ethers containing a monoaza-15-crown-5 ring and two pyrene moieties and their fluorescent properties toward Na⁺, K⁺, Mg²⁺, and Ca²⁺ cations.

Monoaza-15-crown-5 was selected as the main binding site of new fluorescent chemosensors suitable for Ca²⁺, since effective collaboration of the 15-crown-5 ring with an electron-donating sidearm upon complexation with Ca²⁺ was expected based on our previous studies on lariat ethers.8 An electron-donating sidearm was introduced to the C-pivot position of the monoaza-crown ether by the reaction of a key intermediate, 5-bromomethyl-5-methyl-1,4,7,10-tetraoxa-13-azacyclopentadecane, with ethylene glycol or diethylene glycol under basic conditions to give the corresponding C-pivot lariat monoaza-crown ethers, respectively (Scheme S1¹³). The reaction of these compounds with 1-bromomethylpyrene was carried out in THF in the presence of NaH at reflux for 36 and 60 h to afford 1a and 1b in 46% and 32% yield, respectively. The structures (Figure 1) were confirmed by ¹H and ¹³C NMR spectroscopy, mass spectrometry, and elemental analysis. 10

Figure 1. Structures of 1a and 1b; labels a-f denote the corresponding hydrogen atoms.

Figure 2 shows the fluorescence spectra of 1a and 1b at a concentration of $1.0 \times 10^{-6} \,\mathrm{M}$ in CH₃CN in the absence and presence of metal cations. These compounds possess two types of pyrenylmethyl groups, one at the terminal of the electrondonating sidearm of the C-pivot position and the other at the Npivot position of the monoaza-crown ring. It should be noted that the fluorescence of both 1a and 1b based on the pyrene ring was effectively quenched due to the PET from the amino group, in spite of the presence of two pyrene rings for the single amino group. This result showed that both pyrene rings followed the PET mechanism. 7b Upon complexation with a metal cation, the nitrogen lone pair no longer participates in the PET process, causing recovery of the fluorescence. A remarkable increase of monomer emission at 375 nm was observed for 1a upon the addition of Mg²⁺ (16 times) and Ca²⁺ (18 times), without a significant increase of excimer emission at 475 nm (Figure 3). On the other hand, 1b with its longer electron-donating sidearm had an increased excimer emission for Ca²⁺ (12 times) and monomer emission for Mg²⁺ (8 times). Since the increase of excimer emission is attributed to the intramolecular π – π stacking of the two pyrene rings, this finding strongly suggests that the length of the oxyethylene chain on the sidearm plays a crucial role in bringing the two pyrene rings into close proximity. It should be noted that the increase of excimer emission upon the addition of Ca²⁺ observed in fluorophore **1b** was completely in contrast to the decrease of excimer emission in the case of C-pivot lariat crown ethers previously reported.⁵

The stability constants (K) of the complex were evaluated from plots of monomer or excimer emission intensities vs. [metal]/[ligand] by means of a nonlinear least-square curve-fitting method. ¹¹ The curve showed that both ligands formed 1:1 complexes. The $\log K$ values of **1a** and **1b** toward metal cations



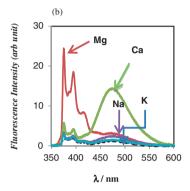


Figure 2. (a) Fluorescence spectra of **1a** $(1 \times 10^{-6} \, \text{M})$ and (b) **1b** $(1 \times 10^{-6} \, \text{M})$ in CH₃CN in the absence (black dotted line) and presence of metal cations $(3.2 \times 10^{-6} \, \text{M})$. Excitation wavelength: 340 nm.

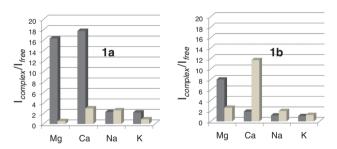


Figure 3. Change of fluorescence intensity of **1a** or **1b** $(1.0 \times 10^{-6} \, \mathrm{M})$ upon addition of metal ions $(3.2 \times 10^{-6} \, \mathrm{M})$ in CH₃CN; dark gray column: monomer emission at 375 nm; pale gray column: excimer emission at 475 nm. Excitation wavelength: 340 nm.

are summarized in Table 1. Both **1a** and **1b** showed a good affinity toward Ca²⁺. This result is reasonably explained by considering the fitness of the cavity size of the ligand and the cation size and the difference in the charge density of alkaline earth metal cation and alkali metal cation.

In order to elucidate the function of the electron-donating sidearm, ^{1}H NMR spectral changes of $\bf{1a}$ and $\bf{1b}$ were examined upon addition of $Ca(ClO_4)_2$ in 9:1 v/v $CD_3CN/CDCl_3$ (Figure S3¹³). Pyrene protons (f) of $\bf{1a}$ and $\bf{1b}$ were observed at 7.95–8.62 ppm in the absence of Ca^{2+} . Upon addition of an equimolar amount of Ca^{2+} , the pyrene protons were shifted upfield to 7.63–8.50 ppm for $\bf{1a}$ and 7.42–8.32 ppm for $\bf{1b}$. A larger upfield shift of the pyrene protons of $\bf{1b}$ compared to

Table 1. The $\log K$ values of **1a** and **1b** for metal cations in CH_3CN

Compound	Mg^{2+}	Ca ²⁺	Na ⁺	K^+
1a	6.7	7.9	5.0	nd
1b	6.3	7.1	5.1	nd

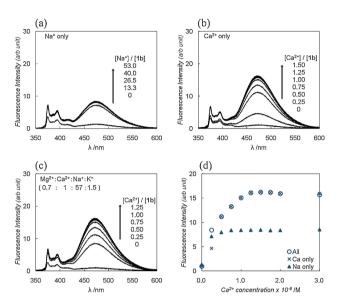


Figure 4. Fluorescence spectral changes of **1b** $(1.0 \times 10^{-6} \, \mathrm{M})$ with different concentrations of alkali metal and alkaline earth metal cations: (a) Na⁺ only, (b) Ca²⁺ only, (c) Mg²⁺, Ca²⁺, Na⁺, K⁺, and (d) fluorescence intensity changes of **1b** at 475 nm $(\bigcirc \mathrm{Mg^{2+}}, \mathrm{Ca^{2+}}, \mathrm{Na^{+}}, \mathrm{K^{+}}; \times \mathrm{Ca^{2+}}$ only; and $\blacktriangle \mathrm{Na^{+}}$ only) in CH₃CN. Excitation wavelength: 340 nm.

those of **1a** provided evidence for the proximity of the two pyrene rings of **1b**. The singlet peak of the methylene protons (e) of **1a** at 5.22 ppm was separated into a pair of doublet and shifted downfield to 5.4 ppm upon addition of Ca^{2+} , but the corresponding peak (e) of **1b** at 5.22 ppm was only shifted downfield to 5.3 ppm and was not split. This result may suggest that the pyrenylmethyl group on the electron-donating sidearm of **1b** moves freely and can easily access another pyrene ring. In the case of the $\mathbf{1a} \cdot Ca^{2+}$ complex, all oxygen atoms of the electron-donating sidearm of **1a** coordinate to Ca^{2+} , and the movement of the pyrenylmethyl group is highly restricted. In addition, the length of the electron-donating sidearm of **1a** may be too short to promote $\pi - \pi$ interaction of the two pyrene rings in the complex with Ca^{2+} .

Next, the selective fluorescent detection of Ca^{2+} using **1b** was examined in the presence of metal cations (Na⁺, K⁺, Mg²⁺, and Ca^{2+}) relevant to living organisms. The concentrations of these metal chlorides were prepared by considering the human serum constitution¹² to be 142×10^{-3} (Na⁺), 4.14×10^{-3} (K⁺), 1.69×10^{-3} (Mg²⁺), and 2.50×10^{-3} M (Ca²⁺) in water, and then the solutions were diluted ten times with CH₃CN to generate the stock solutions (9:1 = CH₃CN:H₂O). The fluorescent spectral changes of **1b** (1.0×10^{-6} M) with different concentrations of metal cations are shown in Figure 4. The addition of Na⁺ or Ca²⁺ increased the excimer emission of pyrene rings at 475 nm (Figures 4a and 4b). However, the

maximum fluorescence intensity with Ca2+ was much higher than that with Na+, and a large excess of metal cations was needed to achieve the maximum fluorescence with Na+. The fluorescence behavior of 1b in the presence of a mixture of Na⁺, K⁺, Mg²⁺, and Ca²⁺ was found to be very similar to that of **1b** in the presence of only Ca²⁺ (Figure 4c). Thus, the fluorescence intensities at 475 nm were plotted as the functions of Ca²⁺ concentration (Figure 4d). After exceeding $0.5 \times 10^{-6} \,\mathrm{M}$ of Ca²⁺ concentration, the plots of the fluorescence intensities for a mixture of metal cations coincided with those for only Ca²⁺ at the same concentrations. This result demonstrates that 1b can selectively detect Ca²⁺ in spite of the disturbance of other metal cations. Based on the graph of Figure 4d, a standard curve prepared by plotting Ca^{2+} concentrations on the x axis and the differences of fluorescence intensities of 1b between the mixed system and the Ca^{2+} single system on the y axis (Figure S4¹³) yielded a good linear relationship. As a result, the concentrations of Ca^{2+} were successfully determined at $0.25-1.25 \times 10^{-6} M$ in the presence of a mixture of Na⁺, K⁺, Mg²⁺, and Ca²⁺.

In summary, two kinds of C-pivot lariat monoaza-crown ethers containing a 15-crown-5 ring and two pyrene moieties on the sidearms (**1a** and **1b**) were prepared and their fluorescent properties toward Na⁺, K⁺, Mg²⁺, and Ca²⁺ were examined. These chemosensors can discriminate among various metal cations by using both ratiometry of the pyrene excimer and monomer emissions and the PET mechanism. Compound **1b** was found to be a highly sensitive and selective chemosensor for Ca²⁺. Continuing molecular design of this type of fluorophore is underway in our laboratory.

References and Notes

- a) G. W. Gokel, W. M. Leevy, M. E. Weber, *Chem. Rev.* 2004, 104, 2723. b) I. Leray, B. Valeur, *Eur. J. Inorg. Chem.* 2009, 3525.
- a) E. A. Weitz, V. C. Pierre, *Chem. Commun.* 2011, 47, 541.
 b) Y. Park, D. C. Apodaca, J. Pullen, R. C. Advincula, *J. Phys. Chem. B* 2010, 114, 13084.
 c) J. Brandel, M. Sairenji, K. Ichikawa, T. Nabeshima, *Chem. Commun.* 2010, 46, 3958.
 d) J. Kim, T. Morozumi, H. Nakamura, *Org. Lett.* 2007, 9, 4419.
 e) A. Minta, R. Y. Tsien, *J. Biol. Chem.* 1989, 264, 19449.
- 3 a) A. P. de Silva, H. Q. N. Gunaratne, T. Gunnlaugsson, A. J. M. Huxley, C. P. McCoy, J. T. Rademacher, T. E. Rice, Chem. Rev. 1997, 97, 1515. b) J. F. Callan, A. P. de Silva, D. C. Magri, Tetrahedron 2005, 61, 8551.
- 4 a) G. W. Gokel, D. M. Dishong, C. J. Diamond, *J. Chem. Soc., Chem. Commun.* 1980, 1053. b) Y. Nakatsuji, T. Nakamura, M. Okahara, D. M. Dishong, G. W. Gokel, *J. Org. Chem.* 1983, 48, 1237. c) D. A. Gustowski, V. J. Gatto, J. Mallen, L. Echegoyen, G. W. Gokel, *J. Org. Chem.* 1987, 52, 5172. d) Y. Nakatsuji, T. Nakamura, M. Yonetani, H. Yuya, M. Okahara, *J. Am. Chem. Soc.* 1988, 110, 531. e) R. Wakita, M. Yonetani, Y. Nakatsuji, M. Okahara, *J. Org. Chem.* 1990, 55, 2752. f) K. Kita, T. Kida, Y. Nakatsuji, I.

- Ikeda, J. Chem. Soc., Perkin Trans. 1 1998, 3857. g) A. A. Abbas, A. H. M. Elwahy, J. Heterocycl. Chem. 2009, 46, 1035.
- 5 a) Y. Nakahara, Y. Matsumi, W. Zhang, T. Kida, Y. Nakatsuji, I. Ikeda, Org. Lett. 2002, 4, 2641. b) Y. Nakahara, T. Kida, Y. Nakatsuji, M. Akashi, J. Org. Chem. 2004, 69, 4403.
- S. Karuppannan, J.-C. Chambron, *Chem.—Asian J.* 2011, 6, 964.
- a) A. P. de Silva, S. A. de Silva, J. Chem. Soc., Chem. Commun. 1986, 1709. b) K. Kubo, N. Kato, T. Sakurai, Bull. Chem. Soc. Jpn. 1997, 70, 3041. c) A. J. Pearson, W. Xiao, J. Org. Chem. 2003, 68, 5369. d) H. He, M. A. Mortellaro, M. J. P. Leiner, S. T. Young, R. J. Fraatz, J. K. Tusa, Anal. Chem. 2003, 75, 549. e) Y. Nakahara, T. Kida, Y. Nakatsuji, M. Akashi, Chem. Commun. 2004, 224. f) Y. Nakahara, T. Kida, Y. Nakatsuji, M. Akashi, Org. Biomol. Chem. 2005, 3, 1787.
- 8 Y. Nakatsuji, M. Muraoka, in *Bottom-up Nanofabrication: Supramolecules, Self-Assemblies, and Organized Films*, ed. by K. Ariga, H. S. Nalwa, American Scientific Publishers, **2009**, Vol. 2 (supramolecules-II), pp. 125–150.
- R. Wakita, M. Tsubakihara, Y. Nakatsuji, M. Okahara, Synthesis 1990, 1011.
- 10 **1a**: 1 H NMR (300 MHz, CDCl₃): δ 8.52 (d, J = 9.3 Hz, 1H), 8.33 (d, J = 9.3 Hz, 1H), 8.19–7.82 (m, 16H), 5.22 (s, 2H), 4.28 (s, 2H), 3.82-3.23 (m, 20H), 2.99-2.74 (m, 4H), 1.19 (s, 3H). 13 C NMR (75 MHz, CDCl₃): δ 133.2, 131.4, 131.2, 131.2, 131.1, 130.8, 130.7, 130.6, 129.8, 129.3, 128.0, 127.6, 127.4, 127.3, 127.3, 126.9, 126.9, 126.9, 125.8, 125.7, 125.1, 124.9, 124.8, 124.8, 124.7, 124.6, 124.4, 124.3, 124.2, 123.5, 76.8, 74.5, 73.8, 71.7, 71.2, 71.1, 70.4, 70.1, 70.0, 69.5, 69.4, 62.2, 59.1, 54.6, 54.4, 18.4. MS(ESI) m/z: 758 [M + Na⁺]. Anal. Calcd for C₄₈H₄₉NO₆: C, 78.34; H, 6.71; N, 1.90%. Found: C, 78.09; H, 6.65; N, 1.84%. **1b**: ¹H NMR (300 MHz, CDCl₃): δ 8.51 (d, J = 9.3 Hz, 1H), 8.34 (d, J = 9.0 Hz, 1H), 8.17-7.90 (m, 16H), 5.22 (s, 2H), 4.27 (s, 2H), 3.72-3.34 (m, 24H), 2.91-2.78 (m, 4H), 1.15 (s, 3H). 13 C NMR (75 MHz, CDCl₃): δ 133.2, 131.3, 131.2, 131.2, 131.1, 130.8, 130.7, 130.6, 129.7, 129.3, 128.0, 127.6, 127.4, 127.3, 127.3, 126.9, 126.9, 126.9, 125.8, 125.7, 125.1, 124.9, 124.8, 124.8, 124.7, 124.6, 124.4, 124.3, 124.2, 123.4, 76.7, 74.4, 73.7, 71.8, 71.1, 71.0, 70.7, 70.5, 70.3, 70.1, 70.0, 69.5, 69.4, 62.2, 59.1, 54.5, 54.4, 18.3. MS(ESI) m/z: 802 [M + Na⁺]. Anal. Calcd for C₅₀H₅₃NO₇: C, 77.00; H, 6.85; N, 1.80%. Found: C, 76.97; H, 6.81; N, 1.67%.
- 11 K. Hirose, J. Inclusion Phenom. Macrocyclic Chem. 2001, 39, 193.
- 12 *Biochemistry Data Book*, ed. by the Japan Biochemical Society, Tokyo Kagaku Dojin, **1979**, pp. 1542–1543.
- 13 Supporting Information is available electronically on the CSJ-Journal Web site, http://www.csj.jp/journals/chem-lett/index.html.